

## 4.3 PHYTOBENTHOS

### 4.3.1 Introduction

Phytobenthos community was included in JDS sampling programme. The main objective was to obtain comparable and reliable data on phytobenthic organisms.

Phytobenthos is the totality of algae living on the surface of substrata in the river bed, thus being mainly autotrophic organisms (Rott, 1991). The ecological niche of phytobenthos algae can be characterized by a long list of environmental variables (hydrology, substratum, light, water chemistry, temperature and other biota) showing river type specific variation ranges. The response of a particular species is determined mainly by species-specific tolerances, such as the range between minimum and optimum requirements for a set of quality criteria of the water and of the environment. Species-inherent properties such as size, morphology of colonies and average life span can be modified fundamentally by properties of the system.

Phytobenthos is normally a well structured community consisting of organisms of a large size spectrum, from a few microns to several centimetres. However, In many phytobenthos studies in rivers, a significant portion of macroalgal species belonging into different taxonomic groups has been neglected. On a long-term scale, phytobenthos communities respond to environmental stress (e.g. abrasion, siltation, instability of substratum, seasonal and horizontal shade pattern, turbidity, hardness, nutrient content, diurnal and seasonal variations, grazing by zoobenthos, fish, shading by riparian vegetation) primarily by changes in species composition.

### 4.3.2 Methodology

#### Sampling procedure

Phytobenthic algae live attached to sediment particles in the uppermost millimeter of the sediment where sun irradiation is available for photosynthesis. For identification of phytobenthos algae, the upper millimeter of the sediment on stones, woods, leaves and other artificial materials in the water course was sampled. The stones, woods or other artificial material were scraped with a blade or tweezers or washed by brush. The microalgae were transferred into glass bottles with river water. Leaves of the trees were pressed and the liquid was added to the sample. Macroscopic filamentous algae were taken by tweezers to a separate bottle. Samples were preserved in formaline (38 %) in the 1:10 ratio immediately after sampling. Samples of phytobenthos were taken together with macrophytes.

#### Identification of phytobenthic organisms

The determination of phytobenthic organisms was carried out to the lowest possible level (mainly to the species level). Some problems occurred in the identification of individual taxa because for their determination the living stage and certain characteristic features are needed to recognize. This was impossible since the samples were conserved. Species from groups of Cyanobacteria, Rhodophyta, Heterocontophyta and Chlorophyta were determined. The available literature records were used for determination. The identification of some problematic taxa was discussed with the experts from Slovakia and the Czech Republic. A total of 223 bottles of samples of the Danube and its tributaries were analysed.

## Quantification

While phytobenthos samples were not taken quantitatively, only very rough estimation of relative abundance was used to obtain semiquantitative data. These data were needed for the calculation of Saprobic Index. The estimation of relative abundance was done on the 1-5 scale according to the following table.

Relative abundance	Occurrence
1	isolated
2	rare
3	abundant
4	very abundant
5	mass

## Calculation of Saprobic Index

The obtained data were processed by the calculation of Saprobic Index. To calculate the Saprobic Index, the Slovak List of Indicators was used.

Regarding the method, the Pantle & Buck formula modified by Zelinka & Marvan (1961) was used:

$$SI = \frac{\sum h_i * s_i * I_i}{\sum h_i * I_i}$$

with  $h_i$  = quantity of species "i" in sample (in case of JDS-phytobenthos samples the "estimation of relative abundance" was used in the scale 1-5)  
 $I_i$  = weight of species "i" in sample.  
 $s_i$  = saprobic index of species "i".

## Quality control/quality assurance

The Slovak National Reference Laboratory was accredited by the Slovak National Accreditation Service on 28th April 1998 (certificate No. 21/1998). The quality of work is maintained by an internal and external control system. The external control is assured by participation in inter-laboratory comparison tests for individual areas of analyses. Biological analyses (microscopic methods) are controlled by the inter-laboratory comparison tests provided by ASLAB Prague. The internal quality assurance system includes uncertainties of measurements, validations of analytical methods, internal audits and a system of internal and external personnel training. The whole system is supported by the Standard Operational Procedures for all analytical activities, metrological regulations and other documents (Standards, Regulations).

The determination of phytobenthic organisms (mainly cyanobacteria, diatoms and filamentous green algae) of some samples was parallelly carried out by specialists from the Institute of Botany of the Slovak Academy of Sciences in Bratislava. All samples of phytobenthos collected during JDS were conserved more intensively again and stored in the collection of the National Reference Laboratory for Water Sector in Slovakia.

### 4.3.3 Results

#### Species diversity

A total of 340 taxa (genera, species, varietal, forma) were identified in the Danube and its tributaries during Joint Danube Survey (Fig. PB-1 and Tab.PB-1).

Four groups of sub-communities were selected. Diatoms were attached to clay, muddy and sandy substratum. Filamentous cyanophytes/cyanobacteria were connected to the fine clay layer (oscillatorium). Some epiphytic cyanobacteria were found growing together with other taxa (Bacillariophyceae, Chlorococcales and Xanthophyceae) on the upper layer of green macro-algae. The macro-algae created a separate group consisting of representatives of Xanthophyceae, Rhodophyta - Bangiophyceae, Chlorophyceae and Zygnematophyceae.

The richest group was Bacillariophyceae (264 taxa). Pennate-species of diatoms predominated, mainly the genera of *Navicula*, *Nitzschia*, *Achnanthes*, *Amphora*, *Cocconeis*, *Cymbella*, *Diatoma*, *Fragillaria*, *Gomphonema*, *Gyrosigma*, *Pinnularia* and *Surirella*.

Among the epipelagic-benthic species, some typical planktonic diatoms of Centrales occurred (e.g. *Aulacoseira*, *Cyclostephanos*, *Cyclotella*, *Skeletonema*, *Stephanodiscus*, *Thalassiosira*). Besides diatoms, green coccal algae (e.g. *Coelastrum*, *Pediastrum*, *Scenedesmus*), flagellates (*Gonium pectorale*) and litoral-liking desmidia (*Cosmarium*, *Closterium*) also appeared. *Melosira varians* occurred in the samples abundantly.

A special ecological niche is made up of filamentous green macro-algae (*Cladophora*, *Oedogonium*, *Stigeoclonium*, *Rhizoclonium*, *Spirogyra*) attached mainly to stony and sandy substrate. Their surface is covered by coccal and filamentous cyanobacteria (*Chroococopsis*, *Clastidium*, *Chamaesiphon*, *Heteroleibleinia*), green coccal algae (*Charatium*), Xanthophyceae (*Characiopsis*) and Bacillariophyceae (*Cocconeis*, *Cymbella*, *Rhoicosphaenia*).

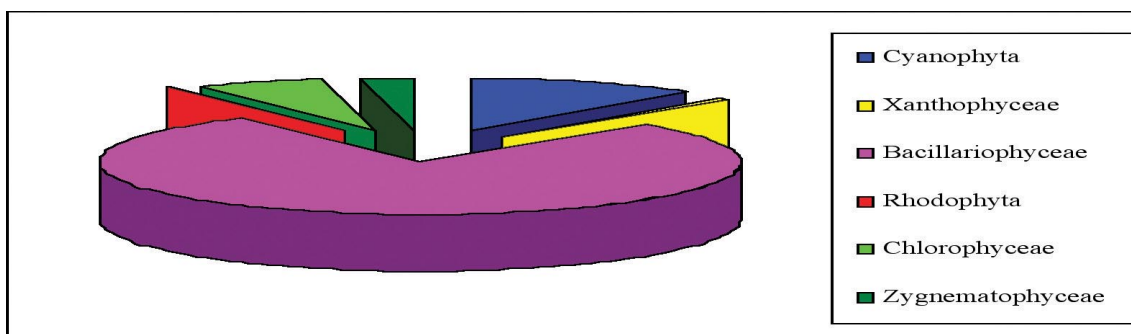


FIGURE PB-1. The percental distribution of numbers of taxa for individual groups of phytobenthos identified during Joint Danube Survey.

A fine surface layer of clay or mud is usually inhabited by filamentous cyanobacteria (*Gleotrichia*, *Plectonema*, *Homeothrix*, *Heteroleibleinia*, *Leibleinia*, *Leptolyngbya*, *Lyngbya*, *Phormidium*).

## Species distribution in the longitudinal section of the Danube and its tributaries

The number of species in the individual stations of the Danube varied in the range of 20-96 and in the tributaries it ranged between 16-109 (Fig. PB-2). The most species were determined in the upper part of the Danube from Neu Ulm to Passau, from Wallsee to Persenbeug and than upstream and downstream of the Drava. Downstream of Kozloduy, the number of identified species decreased significantly. This was the due to the type of substratum (mud and sand). An extremely low number of species was found in the Danube Delta (20-36 taxa).

The number of taxa found in the tributaries varied greatly. Very poor benthic micro-flora was observed in the Jantra where only a few diatoms occurred, probably due to the sandy substratum. The number of species found in the Siret River was also low since the tributary is polluted and so was the number found in the Inn, an alpine tributary with cloudy, glacier waters. In contrast, the highest number of species was found in the Soroksar and Rackeve-Soroksar Danube arms. All other tributaries can be described as medium colonized (49-89 taxa).

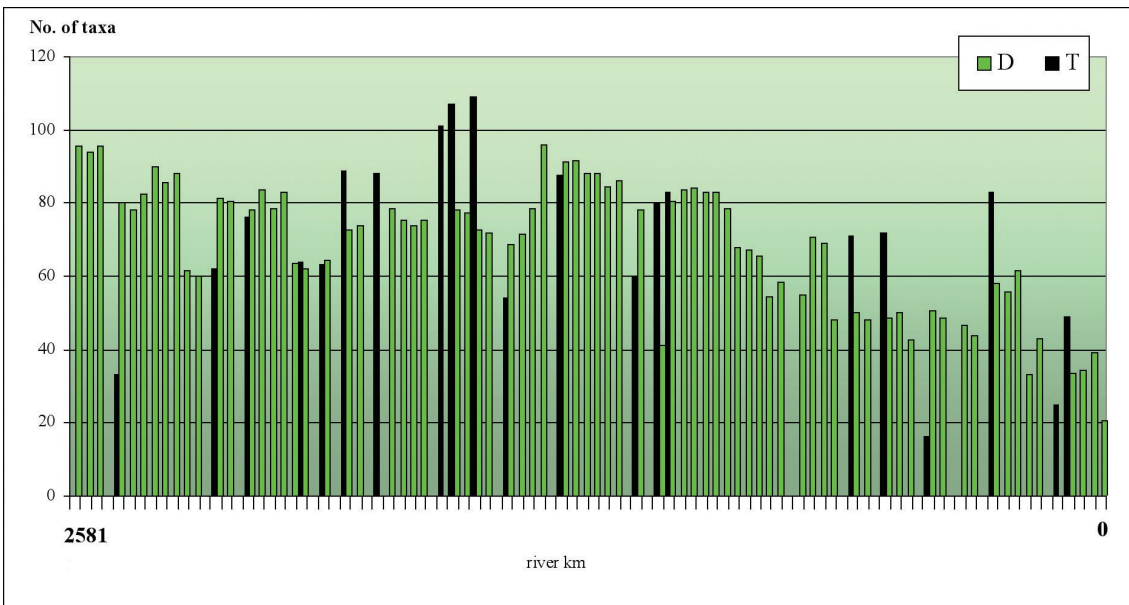


FIGURE PB-2. Number of taxa determined during JDS in the Danube (D) and its tributaries (T).

As for the distribution of individual phytobenthos groups in the Danube, diatoms create the most dominant group. Based on the obtained results, the distribution of individual groups (Cyanophyta, Rhodophyta, Bacillariophyceae, Xanthophyceae and Chlorophyta) along the course of the Danube is presented in Figure PB-3.

Cyanophytes occurred only in the samples that contained a fine layer of mud and were overgrown by filamentic species (0-9 %). Nevertheless, they created only a small portion of microalgae. Like cyanophytes, green algae (Chlorococcales, Ulotrichales, Siphonocladales, Zygnematophyceae) were also rare (0-11 %). Only a few species occurred regularly.

Red algae (Rhodophyta) were represented only by *Bangia arthropurpurea*. This alga was present mainly in the upper part of the Danube. *Cladophora glomerata* was the most frequently occurring filamentous green alga both in the Danube and its tributaries. In a few cases the filaments of this species were completely (95%) covered by *Cocconeis* cells. In some sampling

stations *Spirogyra* appeared forming green hair tufts. Similarly, the genera of *Vaucheria* was characteristic by slime tufts in some stations of the Danube.

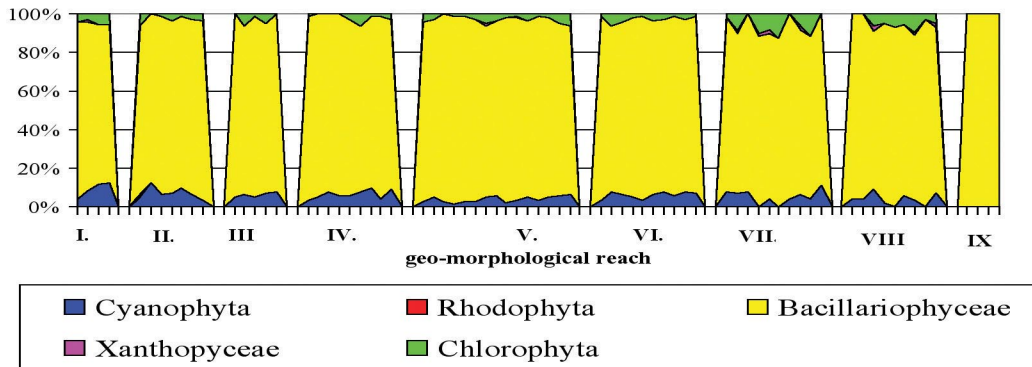


FIGURE PB-3. Relative distribution of taxa numbers of the individual algal groups (Cyanophyta, Rhodophyta, Bacillariophyceae, Xanthophyceae and Chlorophyta) along the nine geo-morphological reaches.

A similar situation (Fig. PB-4) to that in the Danube River itself was observed in the tributaries, where the main group of phytobenthic organisms was made up of diatoms. Only a small percentage of species belonged to cyanophytes and chlorophytes.

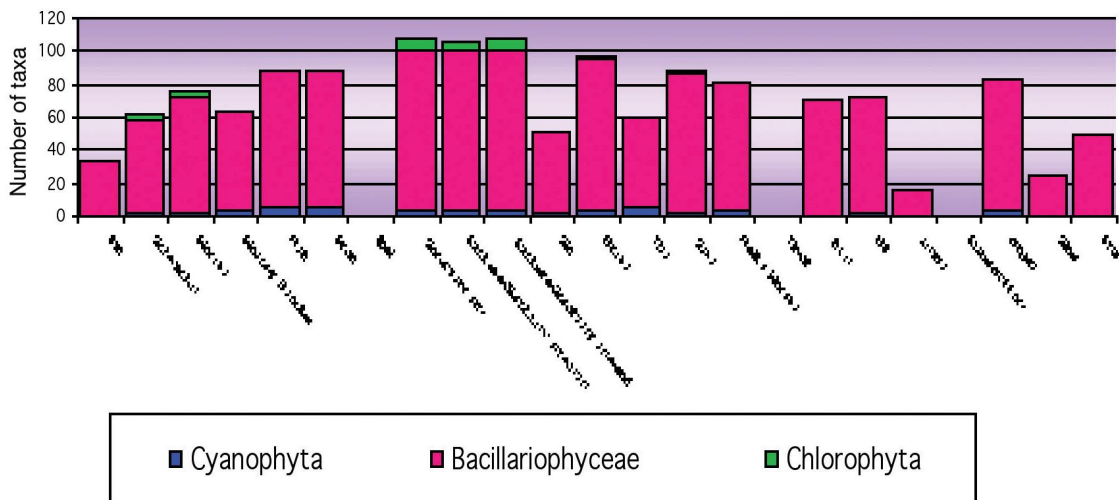


FIGURE PB-4. Distribution (taxa number) of the main algal groups of phytobenthic algae in the Danube tributaries.

TAB.PB-1. List of species found during JDS

**CYANOBACTERIA**

**Chroococcales**

Chroococopsis epiphytica Geitler, 1980  
 Chroococcus sp.  
 Clastidium setigerum Kirchner  
 Chamaesiphon carpaticus Starmach  
 Chamaesiphon incrustans Grunow  
 Chamaesiphon polymorphus Geitler  
 Chamaesiphon polonicus (Rostafinski) Hansgirg  
 Merismopedia punctata Meyen  
 Pleurocapsa aurantiaca Geitler  
 Pleurocapsa minor hansgirg em Geitler

**Nostoccales**

Calothrix parietina Thuret  
 Calothrix sp.

**Oscillatoriales**

Anabaena oscillarioides Bory ex Bornet et Flahault  
 Gleotrichia natans Bory ex Bornet et Flahault  
 Plectonema tomasianum Bornet  
 Nodularia sp.  
 Homeothrix janthina (Bornet et Flahault) Starmach  
 Heteroleibleinia fontana Komárek  
 Heteroleibleinia kuetzingii (Schmidle) Compere  
 Leibleinia epiphytica (Hieronymmus) Anagnostidis & Komárek  
 Leptolyngbya boryana (Gomont) Anagnostidis & Komárek  
 Leptolyngbya faveolarum  
 Lyngbya martensiana Meneghini ex Gomont  
 Lyngbya major Meneghini  
 Oscillatoria limosa Aghard  
 Oscillatoria sancta Kutzing x Gomont  
 Oscillatoria sp.  
 Phormidium autumnale Aghard ex Gomont  
 Phormidium corium Gomont  
 Phormidium chalybeum (Mertens ex Gomont)  
 Anagnostidis & Komárek  
 Phormidium cf. chloroxanthum  
 Phormidium nigrum  
 Phormidium retzii (Aghard) Gomont  
 Phormidium subfuscum  
 Phormidium tagestinum  
 Phormidium tenue  
 Pseudanabaena galeata Bocher  
 Pseudanabaena limnetica (Lemmerman) Komárek  
 Pseudanabaena catenata Lauterborn

**RHODOPHYTA**

Bangiales  
 Bangia arthropurpurea (Roth) Agardh

**HETEROCONTOPHYTA**

**Xanthophyceae**

Characiopsis sp.  
 Vaucheria cf. intermedia  
 Vaucheria sp.

**Bacillariophyceae**

**Centrales**

Actinocyclus normanii (Gregory) Hustedt  
 Aulacoseira ambigua (Grunow) Simonsen

Aulacoseira granulata (Ehrenberg) Simonsen  
 Aulacoseira granulata m. curvata  
 Aulacoseira muzzanensis (Heisler) Krammer  
 Aulacoseira subarctica (O.Muller) Haworth  
 Cyclostephanos delicatus  
 Cyclostephanos dubius (Fricke) Round  
 Cyclostephanos invisitatus (Hohn & Hel.) Thurm. Stoerm. & Haakansen  
 Cyclostephanos sp.  
 Cyclotella atomus Hustedt  
 Cyclotella bodanica var. lemanica (O.Muller) Bachmann  
 Cyclotella distinguenda Hustedt  
 Cyclotella meneghiniana Kutzing  
 Cyclotella ocellata Pantocsek  
 Cyclotella pseudocomensis  
 Cyclotella pseudostelligera Hustedt  
 Cyclotella pseudostelligera/woltereckii Hustedt  
 Cyclotella pseudostelligera/stelligeroides Hustedt  
 Cyclotella quadrijuncta (Schroter) von Keissler  
 Cyclotella stelligera Cleve  
 Cyclotella sp.  
 Cyclotella cf. cyclopuncta Haakansson & Carter  
 Melosira varians Aghard  
 Pleurosira laevis (Ehrenberg) Compere  
 Skeletonema potamos (Weber) Hasle  
 Stephanodiscus hantzschii Grunow  
 Stephanodiscus hantzschii var. tenuis/ binderanus (Hustedt) Haakansson  
 Stephanodiscus neoastrea Haakansson & Hicckel  
 Stephanodiscus cf. parvus  
 Stephanodiscus sp.  
 Thalassiosira incerta  
 Thalassiosira lacustris  
 Thalassiosira pseudonana Hasle & Heimdal  
 Thalassiosira weissflogii (Grunow) Fryxel  
 Thalassiosira cf. duostra  
 Thalassiosira cf. faurii  
 Thalassiosira sp.

**Pennales**

Achnanthes clevei Grunow  
 Achnanthes delicatula (Grunow) Kutzing  
 Achnanthes hungarica (Grunow) Grunow  
 Achnanthes lanceolata (Brebisson) Grunow  
 Achnanthes lanceolata var. rostrata (Oestrup) Hustedt  
 Achnanthes minutissima Kutzing  
 Achnanthes cf. ploenensis Hustedt  
 Achnanthes spp.  
 Amphora cf. inariensis Krammer  
 Amphora libyca Ehrenbergh  
 Amphora montana Kraske  
 Amphora ovalis (Kutzing) Kutzing  
 Amphora pediculus (Kutzing) Grunow  
 Amphora thumensis (Mayer) Cleve-Euler  
 Amphora veneta Kutzing  
 Amphora sp.  
 Anomoeoneis sphaerophora (Ehrenbergh) Pfitzer  
 Asterionella formosa Hassal  
 Caloneis amphisbaena (Bory) Cleve  
 Caloneis bacillum (Grunow) Cleve

*Caloneis permagna*  
*Caloneis silicula* (Ehrenbergh) Cleve  
*Caloneis schumanniana* (Grunow) Cleve  
*Caloneis* sp.  
*Campylodiscus* sp.  
*Cocconeis pediculus* Ehrenbergh  
*Cocconeis placentula* Ehrenbergh  
*Cocconeis placentula* var.1  
*Cocconeis* sp.  
*Cymatopleura elliptica* (Brebisson) W.Smith  
*Cymatopleura solea* (Brebisson) W.Smith  
*Cymbella amphicephala* Nageli  
*Cymbella* cf. *affinis* Kutzing  
*Cymbella caespitosa* (Kutzing) Brun  
*Cymbella cistula* (Ehrenbergh) Kirchner  
*Cymbella* cf. *delicatula* Kutzing  
*Cymbella ehrenbergii* Kutzing  
*Cymbella helvetica* Kutzing  
*Cymbella helmckeii/lanceolata* (Ehrenbergh) Kirchner  
*Cymbella lanceolata* (Brebisson) W.Smith  
*Cymbella leptoceros* (Ehrenbergh) Kutzing  
*Cymbella* cf. *mesiana* Chalnoky  
*Cymbella microcephala* Grunow  
*Cymbella minuta* Hilse  
*Cymbella prostrata* (Berkeley) Cleve  
*Cymbella silesiaca* Bleisch  
*Cymbella sinuata* Gregory  
*Cymbella* cf. *turgidula*  
*Cymbella tumida* (Brebisson) van Heurck  
*Cymbella* cf. *tumidula* Grunow  
*Cymbella* sp.  
*Denticula tenuis* Kutzing  
*Diatoma ehrenbergii* Kutzing  
*Diatoma mesodon* (Ehrenbergh) Kutzing  
*Diatoma moniliformis* Kutzing  
*Diatoma tenuis* Aghardh  
*Diatoma vulgare* Bory  
*Didymosphenia geminata* (Lyngb) M.Schmidt  
*Diploneis* cf. *elliptica* (Kutzing) Cleve  
*Diploneis* cf. *modica*  
*Diploneis oblongella* (Nageli) Cleve-Euler  
*Diploneis* sp.  
*Epithemia* sp.  
*Epithemia* cf. *sorex* Kutzing  
*Eunotia bilunaris* (Ehrenbergh) Millis  
*Eunotia soleirolii* (Kutzing) Rabenhost  
*Fragilaria arcus* (Ehrenbergh) Cleve  
*Fragilaria berlinensis*  
*Fragilaria* cf. *bicapitata* A.Mayer  
*Fragilaria bidens* Heiberg  
*Fragilaria brevistriata* Grunow  
*Fragilaria capucina* var. *capitellata* Krammer & Lange-Bertalot  
*Fragilaria capucina* var. „*gracilis*“  
*Fragilaria capucina* var. *mesolepta* (Rabenhorst) Rabenhorst  
*Fragilaria capucina* var. *perminuta*  
*Fragilaria capucina* var. *radians* (Rabenhorst) Rabenhorst  
*Fragilaria capucina* var. *vaucheriae* (Kutzing) Krammer & Lange-Bertalot  
*Fragilaria construens* (Ehrenbergh) Hustedt  
*Fragilaria construens* f. *binodis* (Ehrenbergh) Hustedt  
*Fragilaria crotonensis* Kitton  
*Fragilaria elliptica*  
*Fragilaria fasciculata*  
*Fragilaria* aff. *lapponica*  
*Fragilaria leptostauron*  
*Fragilaria montana*  
*Fragilaria parasitica*  
*Fragilaria parasitica* var. *subconstricta*  
*Fragilaria pinnata*  
*Fragilaria* cf. *tenera*  
*Fragilaria ulna* var. *acus*  
*Fragilaria ulna*  
*Fragilaria ulna* var. *oxyrhynchus*  
*Fragilaria* sp.  
*Frustulia vulgaris*  
*Gomphonema angustatum*  
*Gomphonema gracile*  
*Gomphonema minutum*  
*Gomphonema olivaceum*  
*Gomphonema parvulum*  
*Gomphonema tergestinum*  
*Gomphonema truncatum*  
*Gyrosigma acuminatum*  
*Gyrosigma attenuatum*  
*Gyrosigma parkerii*  
*Gyrosigma scalproides*  
*Hantzschia amphioxys*  
*Meridion circulare*  
*Navicula accomoda*  
*Navicula* cf. *atomus*  
*Navicula bacillum*  
*Navicula capitata*  
*Navicula capitata* var. *lueneburgensis*  
*Navicula capitatoradiata*  
*Navicula* cf. *capitatoradiata*  
*Navicula citrus*  
*Navicula* cf. *constans*  
*Navicula* cf. *costulata*  
*Navicula cryptocephala*  
*Navicula cryptotenella/menisculus*  
*Navicula cuspidata*  
*Navicula* cf. *decussis*  
*Navicula* cf. *erifuga*  
*Navicula* cf. *exigua*  
*Navicula gastrum*  
*Navicula goeppertiana*  
*Navicula gregaria*  
*Navicula laevissima*  
*Navicula lanceolata*  
*Navicula lenzii*  
*Navicula libonensis*  
*Navicula menisculus*  
*Navicula microrhombus*  
*Navicula minuscula*  
*Navicula* cf. *mutica*  
*Navicula oblonga*  
*Navicula protracta*  
*Navicula* aff. *pseudanglica*  
*Navicula pupula*  
*Navicula pupula* var. *mutata*  
*Navicula pygmaea*  
*Navicula radiosa*

Navicula recens  
 Navicula reinhardtii  
 Navicula cf. rhyngocephala  
 Navicula cf. schroeteri  
 Navicula splendidula  
 Navicula slesvicensis  
 Navicula cf. subhamulata  
 Navicula subminuscula  
 Navicula aff. tenera  
 Navicula tripunctata  
 Navicula trivialis  
 Navicula veneta  
 Navicula viridula var. rostellata  
 Navicula viridula  
 Neidium ampliatum  
 Neidium binodis  
 Neidium dubium  
 Nitzschia acicularis  
 Nitzschia cf. amphibia  
 Nitzschia angustata  
 Nitzschia cf. angustatula  
 Nitzschia brevissima  
 Nitzschia calida  
 Nitzschia capitellata  
 Nitzschia cf. clausii  
 Nitzschia constricta  
 Nitzschia dissipata  
 Nitzschia dubia  
 Nitzschia aff. flexa  
 Nitzschia fonticola  
 Nitzschia frustulum  
 Nitzschia fruticosa  
 Nitzschia gracilis  
 Nitzschia graciliformis  
 Nitzschia heufleriana  
 Nitzschia hungarica  
 Nitzschia inconspicua  
 Nitzschia intermedia  
 Nitzschia levidensis  
 Nitzschia linearis  
 Nitzschia microcephala  
 Nitzschia palea  
 Nitzschia paleacea  
 Nitzschia cf. plana  
 Nitzschia recta  
 Nitzschia sigmoidea  
 Nitzschia sinuata var. delognei  
 Nitzschia sociabilis  
 Nitzschia cf. subacicularis  
 Nitzschia umbonata  
 Nitzschia cf. wuellerstorffii  
 Nitzschia lanceola var. minutula  
 Nitzschia hantzschiana  
 Nitzschia cf. tubicola  
 cf. Simonsenia delognei  
 Nitzschia sp.  
 Pinnularia appendiculata  
 Pinnularia borealis  
 Pinnularia gibba  
 Pinnularia cf. maior  
 Pinnularia microstauron var. brebissonii  
 Pinnularia cf. neomajor

Pinnularia subcapitata  
 Pinnularia cf. viridis  
 Rhoicosphenia abbreviata  
 Stauroneis lundii  
 Stauroneis phoenicenteron  
 Stauroneis cf. producta  
 Stauroneis smithii  
 Surirella angusta  
 Surirella bifrons  
 Surirella cf. biseriata  
 Surirella brebissonii var. kuetzingii  
 Surirella aff. elegans  
 Surirella gracilis  
 Surirella linearis  
 Surirella linearis var. helvetica  
 Surirella minuta  
 Surirella ovalis  
 Surirella cf. splendida  
 Surirella tenera  
 Surirella cf. crumena  
 Surirella sp.  
 Tabellaria flocculosa  
 Tabellaria sp.  
**Chlorophyceae**  
**Volvocales**  
 Gonium pectorale  
**Chlorococcales**  
 Charatium acuminatum  
 Characium angustum  
 Charatium sp.  
 Coelastrum astroideum  
 Pediastrum boryanum  
 Scenedesmus acuminatus  
 Scenedesmus communis  
 Scenedesmus obliquus  
 Scenedesmus intermedius  
 Scenedesmus brasiliensis  
 Scenedesmus opoliensis  
**Ulotrichales**  
 Stigeoclonium tenue  
 Ulothrix tenerrima  
 Binuclearia sp.  
**Siphonocladales**  
 Cladophora glomerata  
 Cladophora sp.  
 Oedogonium itzigsohnii  
 Oedogonium cf. sociale  
 Oedogonium sp.  
 Rhizoclonium hieroglyphicum  
 Rhizoclonium sp.  
**Zygnematophyceae**  
 Cosmarium botrytis  
 Cosmarium leave  
 Cosmarium sp.  
 Closterium moniliferum  
 Closterium praelongatum var. breave  
 Closterium calosporum  
 Closterium closteroides  
 Closterium sp.  
 Spyrogyra maxima (Hass.) Wittr.  
 Spyrogyra sp.



## Saprobity

Based on species diversity and relative abundance of phytobenthos, the Saprobic Index (SI) of the Danube and its tributaries was calculated at each station. The values ranged between 1.77 – 2.11 in the Danube and between 1.80 – 2.11 in the tributaries. (see Annex - Phytobenthos). This phytobenthic results characterise a beta-mesosaprobic status for all JDS samplings sites. The highest values were found in the Sió and the Jantra tributaries, and other tributaries are comparable to the Danube from the point of view of phytobenthos SI. Of all stations along the Danube, those in the Delta (Reni Chilia arm, Vilkova Chilia arm) reported the highest saprobic value. Saprobic indices calculated for the left and right banks of the sampling sites showed only minimal differences (0-0.06 SI values). There was only a very slight increase in SI values in the Danube section at river km 1800 – 1100 and downstream of river km 641. However, in the longitudinal profile of the Danube the differences in the Saprobic Indices were low and therefore insignificant. The only rough estimation of relative abundance and the preservation of the samples have to be taken into account in evaluating the saprobic status based on phytobenthos.

Calculating the Saprobic Index for the phytobenthos community in the Danube according to the method used proved to be less significant than the saprobic evaluation based on macrozoobenthos. This might be due to the fact that, on the one hand, primary producers such as phytobenthic algae should be used for assessing the trophic status, but compared to zoobenthos they are not the best indicators for the biological assessment of organic pollution (saprobity). On the other hand, bearing in mind that diatoms make up the main group of phytobenthos community in the Danube and its tributaries, it would be advisable to use additional diatom indices in future (e.g. Diatom Index by Descy and Coste, Generic Diatom Index, Trophic Diatom Index, Diatom-Analyses according to Lange-Bertalot).

## Comparison of the nine geo-morphological reaches

For the purpose of JDS, the course of the Danube was divided into nine reaches based on geo-morphological features and potential anthropogenic impacts caused by large impoundments and/or significant point sources of pollution represented by big settlements (see Chapter 3). As shown in Fig. PB-5, the highest number of identified taxa was found in Reach 1. The last three reaches show a lower number of taxa due to the type of substratum at individual stations. It is evident that the last reach was characterised by diatoms, while in others at least a small number of Chlorophyta and Cyanophyta appeared. On the basis of the results, it is clear that species diversity in the lower section of the Danube decreases compared to the middle and upper section of the River.

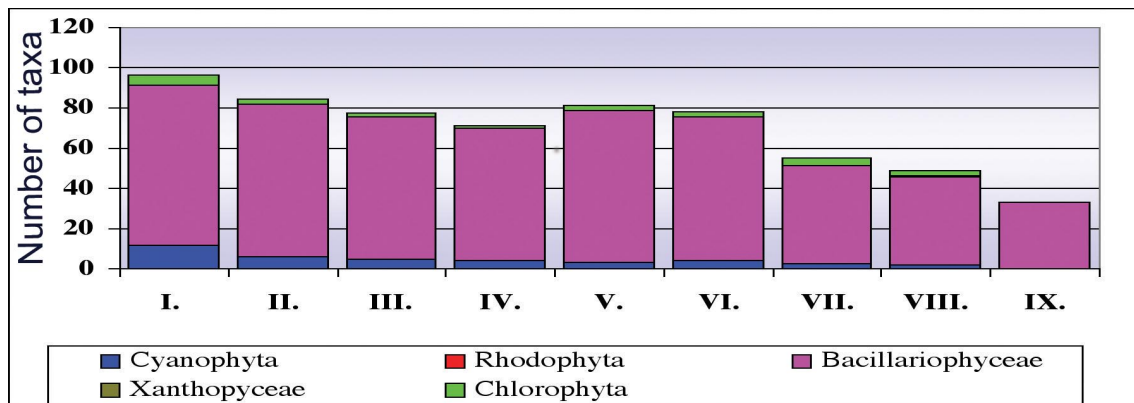


FIGURE PB-5. Number of taxa determined along the Danube distributed according to the nine geo-morphological Reaches.

#### 4.3.4 Summary and Conclusions

A total of 340 taxa of phytobenthos were identified in the Danube, its side-arms and tributaries during the 2001 Joint Danube Survey.

Four groups of sub-communities were selected. Diatoms were attached to clay, muddy and sandy substratum. Filamentous cyanobacteria were connected to the fine clay layer (oscillatorietum). Some epiphytic cyanobacteria were found growing together with other taxa (Bacillariophyceae, Chlorococcales and Xanthophyceae) on the upper layer of green macroalgae. Macro-algae created a separate group consisting of representatives of Xanthophyceae, Rhodophyta - Bangiophyceae, Chlorophyceae and Zygnematophyceae.

The richest group was Bacillariophyceae (264 taxa). Pennate-species of diatoms predominated, mainly the genera of *Navicula*, *Nitzschia*, *Achnanthes*, *Amphora*, *Cocconeis*, *Cymbella*, *Diatoma*, *Fragillaria*, *Gomphonema*, *Gyrosigma*, *Pinnularia* and *Surirella*.

The number of species identified at the individual sampling sites varied in the range of 20-96 in the Danube and between 16 and 109 in the tributaries. Downstream of Koszloduy, the number of phytobenthic species decreased significantly. This seemed to be due to the type of substratum (mud and sand). Extremely low numbers of species were found in the Delta (20-39 taxa).

Saprobic indices calculated by the use of phytobenthos data did not produce a differentiated picture of organic pollution. Therefore, it is recommended to further develop the indication values of phytobenthic organisms or use other indices for the assessment of phytobenthic communities, such as trophic indices, diatom indices, etc.

For future monitoring and repetition of JDS it is also strongly recommended that phytobenthos samples should be analysed directly on board to allow the ratio between individual groups and dominances to be identified and to facilitate the identification of species according to their special characteristic features in the living stage. This would provide a better overview about the status of phytobenthic community,

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